

SELECTION OF MOLECULAR APTAMERS FOR IDENTIFICATION OF LIVE CELLS OF *RALSTONIA SOLANACEARUM*: A NEW POTENTIAL METHOD IN PLANT PATHOLOGY

Patrice Champoiseau, University of Florida

2009 APS ANNUAL MEETING

August 1-5, 2009 ♦ Portland, Oregon



Outlines

- Introduction
- Objective
- Method & protocols
- Results
- Conclusion & perspectives
- Acknowledgements

Introduction

- *R. solanacearum* causes bacterial wilts on a wide range of planted crops, ornamentals and weeds worldwide.

- “Species complex”.

 - 5 races

 - 5 biovars

 - 4 phlotypes

- Race 3 biovar 2 (R3bv2).

 - World wide distribution except US and Canada

 - Major disease of potato

 - Introduction in the US in latently infected geranium cuttings

 - Strictest biosecurity regulations



 - Critical need for unambiguous identification of R3bv2.**

 - Development of fast, sensitive, and specific detection methods.**

Objective

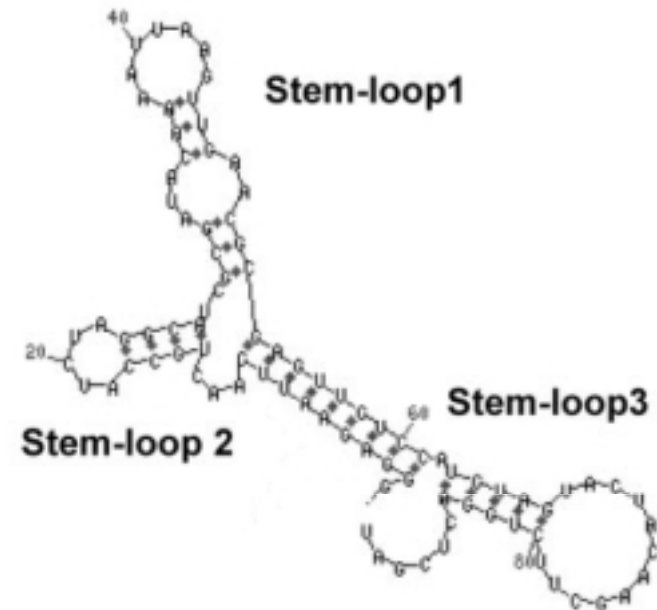
Evaluate the effectiveness of cell-SELEX* to produce molecular aptamers for differentiation of strains of *R. solanacearum* (detection of live cells).

*System Evolution of Ligands by EXponential enrichment

Method & protocols

■ Molecular aptamers.

- Single-stranded oligonucleotides.
- Bind with high affinity to their target.
- Detection of whole cells.
- Small molecules, chemically stable.
- Molecular probes for cancer studies.
- Blocking molecules: reduced virulence in *M. tuberculosis*.



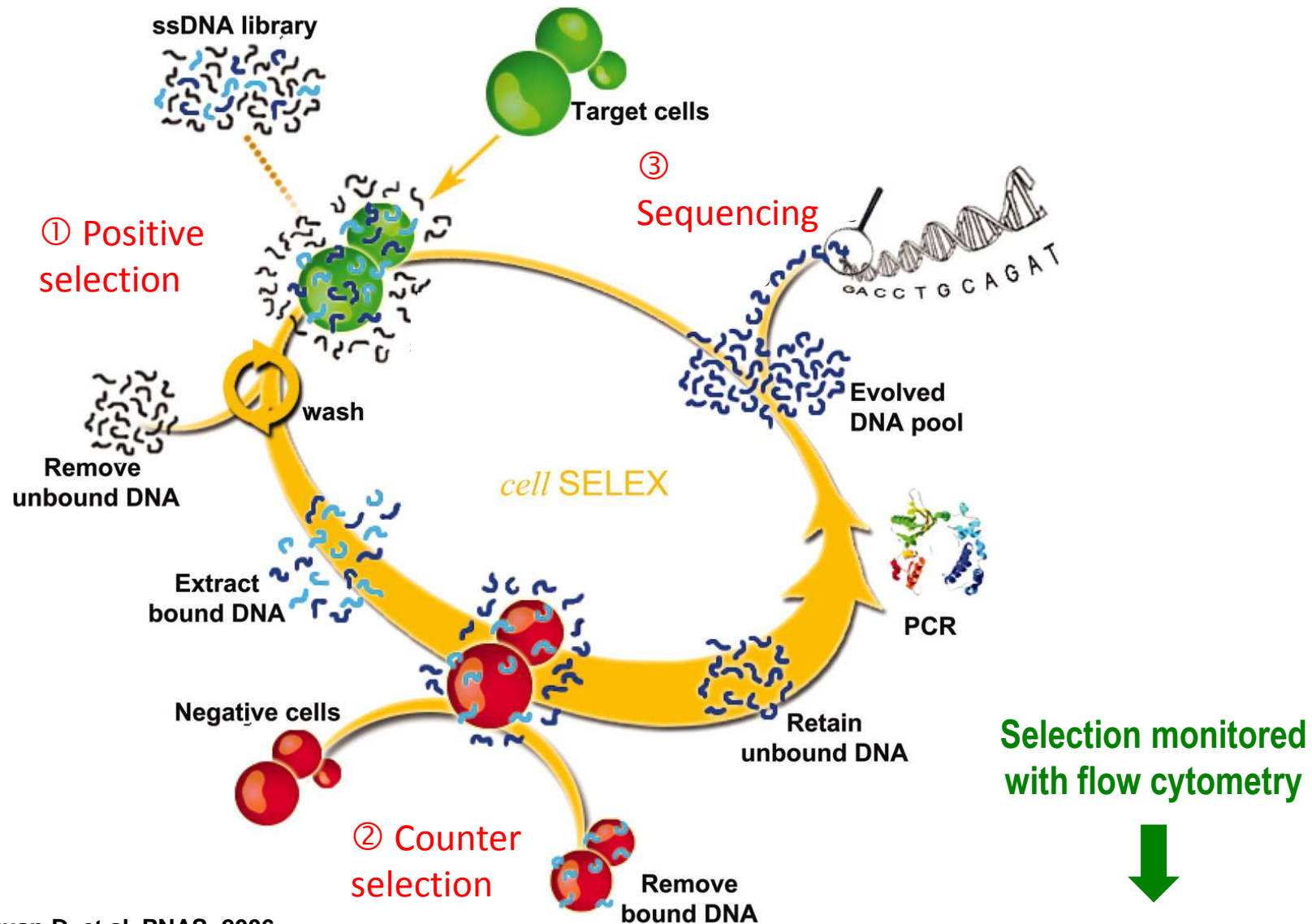
Example of an RNA Aptamer

■ Cell-SELEX.

Repeated competitive binding from an initial DNA or RNA library (random sequences) to the target molecules.

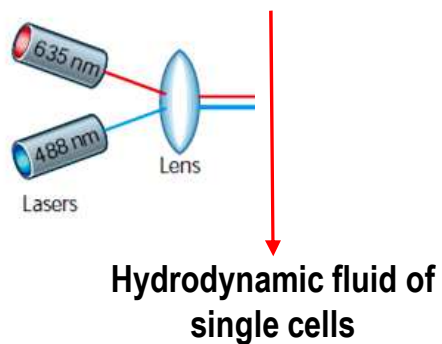
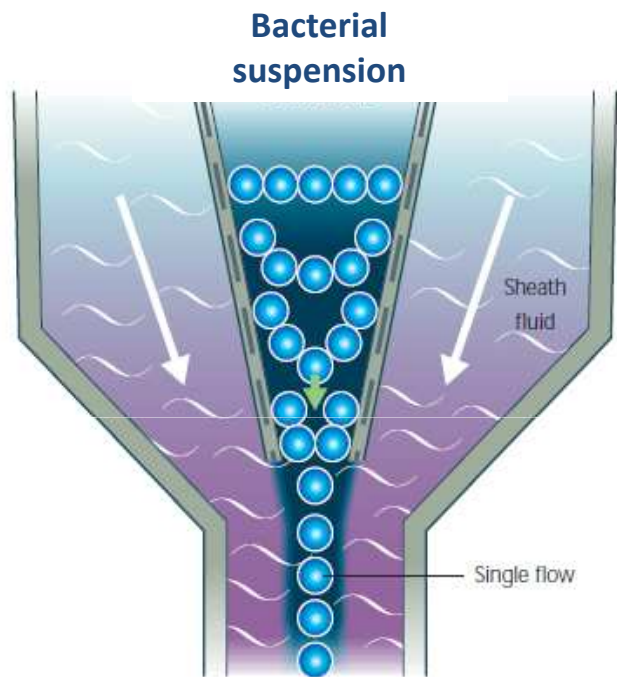
Method & protocols

■ Schematic representation of cell-SELEX.

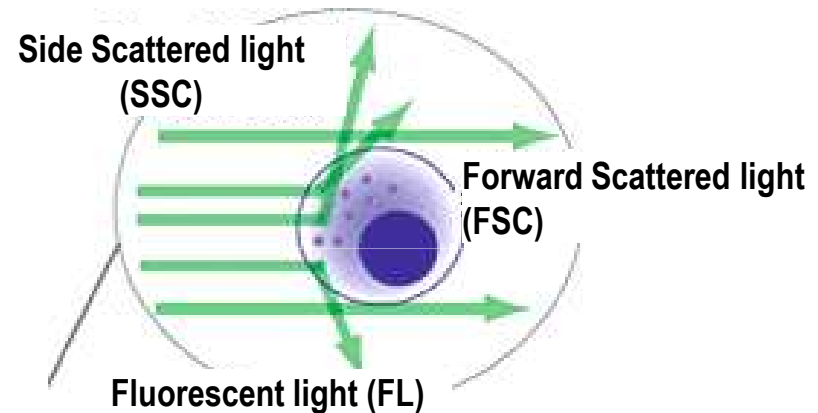


Method & protocols

■ Principles of flow-cytometry



Laser detection system



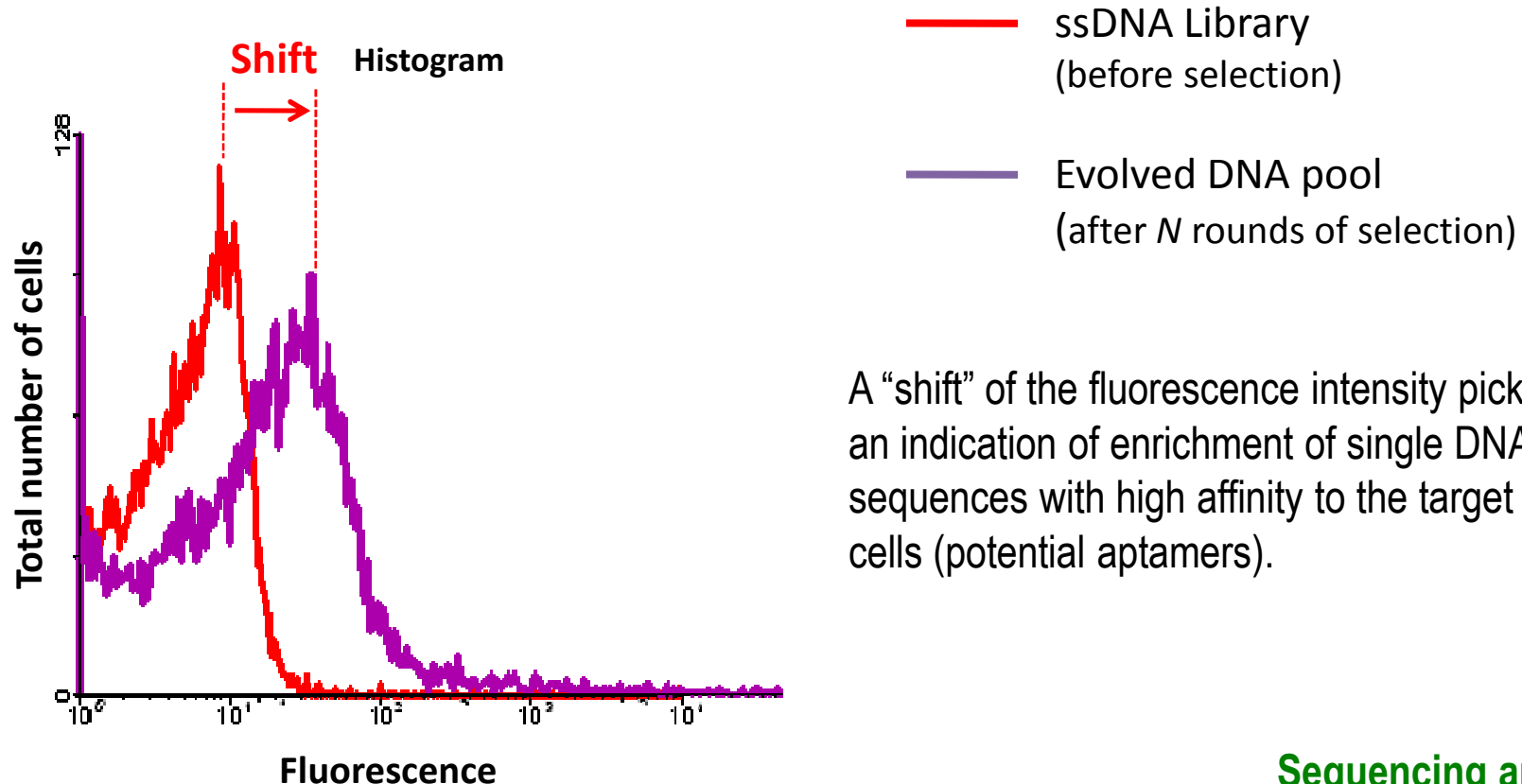
FSC = Indication of particle's size.

SSC = Information about the outer composition of a cell.

FL = Fluorescence detector (1 to 4).

Method & protocols

■ Flow-cytometry data for cell-SELEX



A “shift” of the fluorescence intensity pick is an indication of enrichment of single DNA sequences with high affinity to the target cells (potential aptamers).

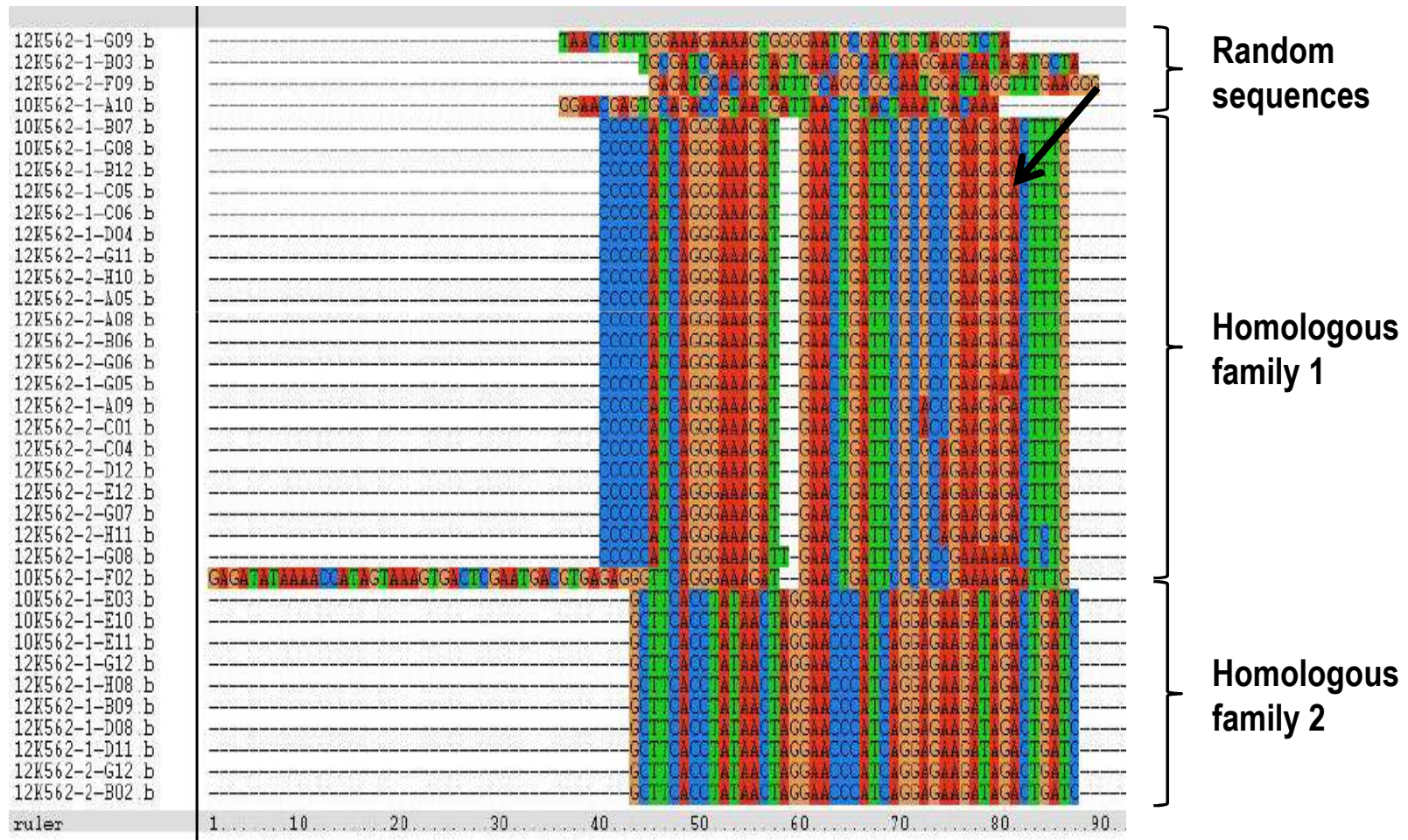
Sequencing and analysis



Method & protocols

■ Sequence analysis

From around 10^{15} unique random sequences



Method & protocols

■ Bacterial strains (cells)

UW485: Race 3 biovar 2T

Rs5: Race 1 biovar 1 (Florida)

■ Cell growth conditions

Liquid medium: MMP

Phosphate-based buffer

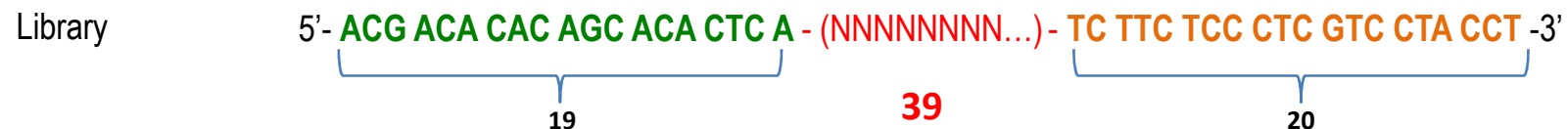
36 hours growth

■ ssDNA Library (78 nucleotides)

Library

5'- **ACG ACA CAC AGC ACA CTC A** - (NNNNNNNN...) - **TC TTC TCC CTC GTC CTA CCT** -3'

19 39 20



Carboxyfluorescein

Forward Primer  5'- **ACG ACA CAC AGC ACA CTC A** -3' → Fluorescence

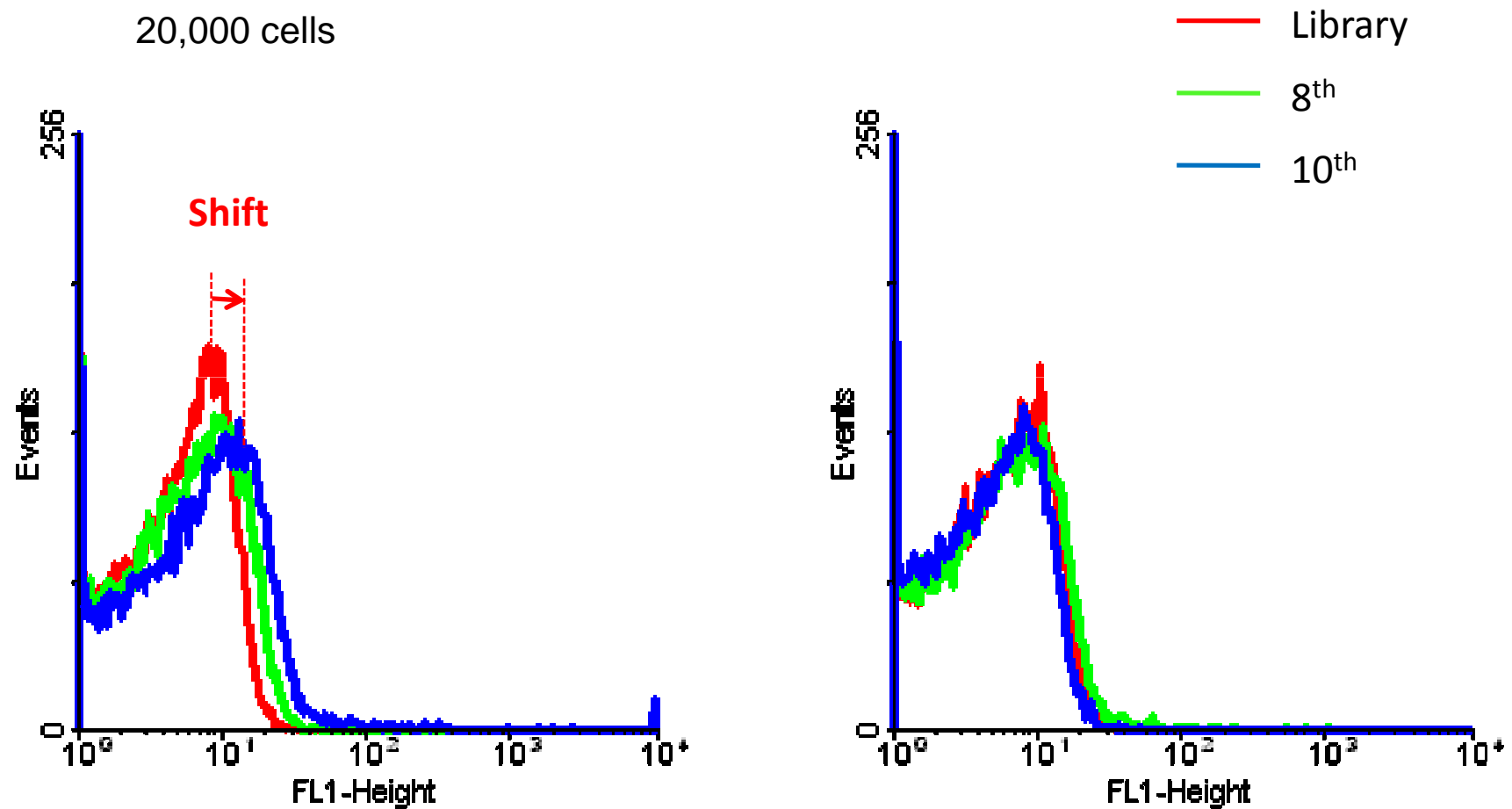
Reverse Primer  5'- **AGG TAG GAC GAG GGA GAA GA** -3' → dsDNA (PCR product) → ssDNA

Biotin

Results

■ Flow-cytometry histograms (10 rounds)

20,000 cells



Target cells (UW485)

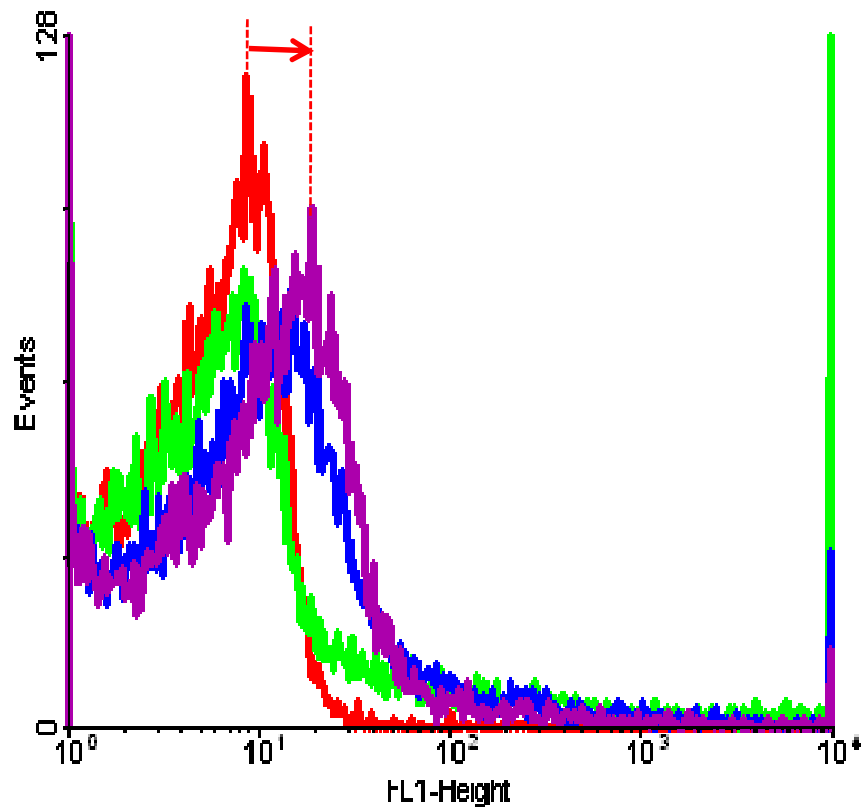
Negative cells (Rs5)

Results

■ Flow-cytometry histograms (14 rounds)

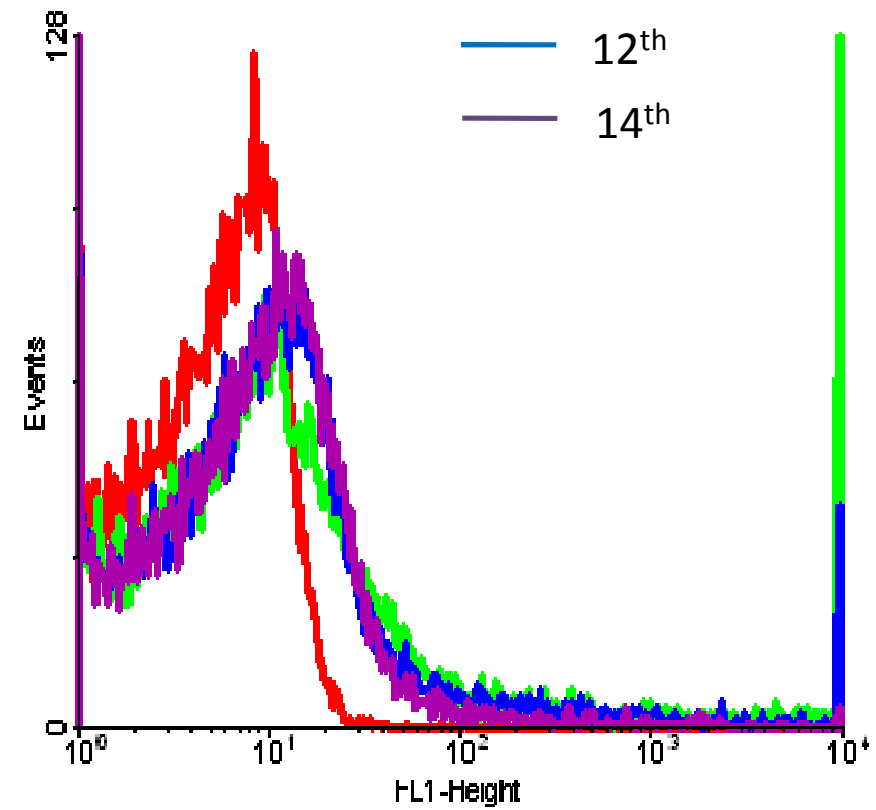
20,000 cells

Shift



Target cells (UW485)

— Library
— 11th
— 12th
— 14th

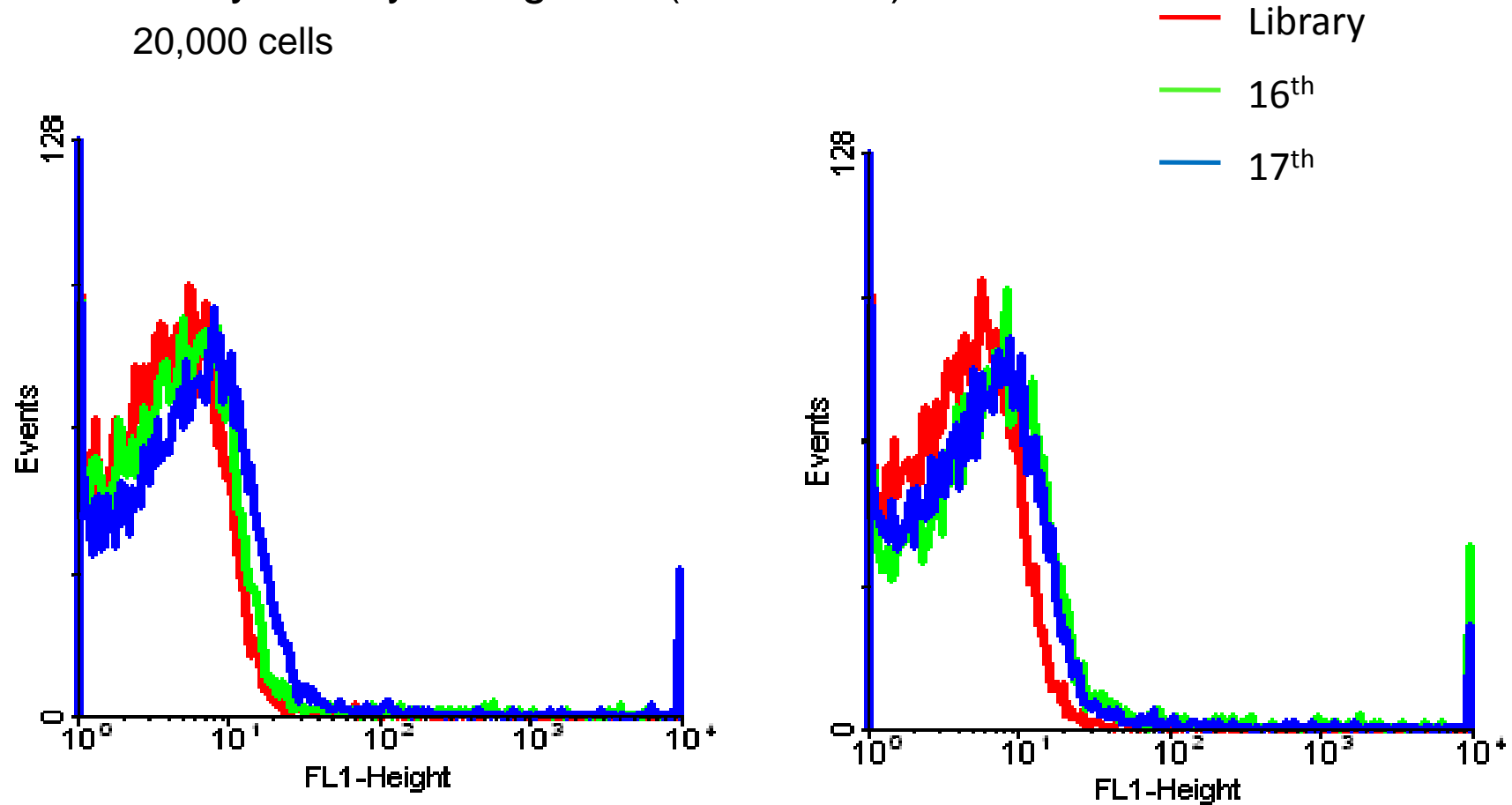


Negative cells (Rs5)

Results

■ Flow-cytometry histograms (17 rounds)

20,000 cells



Target cells (UW485)

Negative cells (Rs5)

Results

■ Sequencing of evolved DNA pools (454)

SP10

353 repeated sequences (out of 3263)

One homologous family : **12 repeated sequences (0.4% total)**

SP16-17

769 repeated sequences (out of 8044)

Several homologous family : **22 repeated sequences (0.3% total)**

Only one conserved DNA family

For Primer – **cagggctcgagatgtttgcatctgttgccgttgga** - Rev Primer

Conclusion and perspectives

■ Conclusion

- Efficiency of flow-cytometry to differentiate cells of *R. solanacearum* after 10 rounds of selection.
- Enrichment of DNA sequences with cell-SELEX.
- Low enrichment due to variability in target molecules (could be EPS ?).
- Amplification of potential aptamers not shown.

■ Perspective

- **Increase the number of repeated sequences.**
More rounds of selection.
- **Test the affinity of the repeated sequences.**
Amplification and testing these sequences as potential aptamers.
- **Control the specificity of the repeated sequences.**
Test with target cells and heterologous bacteria.

Acknowledgements

- Team members.

Caitilyn Allen, Department of Plant Pathology, University of Madison-Wisconsin.

Jeffrey B. Jones, Department of Plant Pathology, University of Florida.

- Team collaborators.

Weihong Tan, Department of Chemistry, University of Florida.

Kwame Sefah, Department of Chemistry, University of Florida.

- Financial support.

National Research Initiative (NRI) program of the USDA Cooperative State Research, Education, and Extension Services (CSREES).

